

Strategies to reduce the impact of heat stress on fertility of cows

Peter J. Hansen

Department of Animal Sciences, University of Florida

The Problem of Infertility in Dairy Cattle Caused by Heat Stress

Heat stress has two major consequences for the physiology of the cow that reduce its probability of becoming pregnant. First, changes in cow behavior (for example, reduced walking time; López-Gatiús et al., 2005) and reduced circulating concentrations of estradiol-17 β (Gilad et al., 1993) caused by heat stress reduce ability to detect estrus. On one dairy in Florida, only about 18-24% of estruses in hot months were detected by herdsman while 45-56% of estrus periods were detected in cool months (Thatcher and Collier, 1986). Secondly, there is a large reduction in fertility. In lactating dairy cows, pregnancy rates per insemination in the summer can be as low as 10-20% (Hansen and Aréchiga, 1999). Fertility is reduced because heat stress can damage both the oocyte and early embryo (Hansen, 2013). The oocyte can be compromised by heat stress as early as 105 days before ovulation (Torres-Júnior et al., 2008) and as late as the peri-ovulatory period (Putney et al., 1989). The early embryo is also initially sensitive to heat stress but quickly becomes resistant so that heat stress on day 1 after estrus reduced embryonic development whereas heat stress at day 3 had no effect (Ealy et al., 1993).

Heat stress is largely a problem of the lactating dairy cow. In Florida, conception rates in Holsteins declined in the summer for lactating cows but not for non-lactating heifers (Badinga et al., 1985). The lactating cow is very susceptible to heat stress because the increased amounts of heat produced as a result of lactation makes it difficult to regulate body temperature during heat stress. Hyperthermia in lactating cows can occur at air temperatures as low as 25-29°C (Berman et al., 1985; Dikmen and Hansen, 2009). Beef cattle can also be affected by heat stress, particularly in feedlot situations (Mitlöhner et al., 2002), but several factors mitigate against large effects on reproduction in beef cattle. These factors include the existence of beef breeds that are genetically resistant to heat stress (Gaughan et al., 2010) and seasonal breeding patterns that ensure that cows are not bred at the warmest time of year.

The most common approach to reduce effects of heat stress on dairy cattle is to provide housing that minimizes heat stress. Incorporation of features such as shade structures, fans, and sprinklers, misters or foggers can be seen in many dairies in hot regions of the world. While very important, cooling cows does not totally prevent reduced reproductive function during heat stress. In Florida, for example, seasonal variation in pregnancy rate persisted in a herd where cows were cooled with sprinklers and fans (Hansen and Aréchiga, 1999). In Israel, pregnancy rate per insemination in intensively-cooled herds was 19% in summer vs 39% in winter for high-producing herds and 25% in summer vs 40% in winter for low-producing herds (Flamenbaum and Ezra, 2006).

In this paper, some other approaches for reducing effects of heat stress on reproductive function will be discussed. These approaches are based on altering the antioxidant status of the cow, use of hormones to manipulate reproductive processes, and incorporation of embryo transfer to bypass effects of heat stress on the oocyte and embryo.

Administration of Antioxidants to Protect Embryos from Free Radicals Induced by Heat Shock

One of the reasons why heat stress damages the oocyte and embryo may reside in an increase in production of free radical molecules at elevated body temperature. Exposure of maturing oocytes (Nabenishi et al., 2012) and early cleavage-stage embryos to elevated temperature (Sakatani et al., 2004) increases the production of free radicals. In culture, some antioxidants [anthocyananin (Sakatani et al., 2007) and

dithiothreitol (de Castro e Paula et al., 2008)] but not all [vitamin E (Paula-Lopes et al., 2003a), glutathione (Ealy et al., 1995), and glutathione ester (Ealy et al., 1995)] reduced the effects of elevated temperature on development of embryos. The possible involvement of free radicals in effects of heat stress on the oocyte and embryo suggests it might be possible to improve fertility by increasing antioxidant concentrations in cells or the blood. There are two papers that suggest that this might be possible.

In the first paper, Aréchiga et al. (1998) found that feeding cows supplemental β -carotene at a rate of 400 mg/d for at least 90 days from about Day 15 after calving increased the proportion of cows that were pregnant at 90 d postpartum during the summer but not during the winter (Table 1). There was no effect of treatment on pregnancy rate at first service so the supplemental β -carotene either improved fertility after first service or estrus detection rate.

Table 1. Effect of supplemental feeding of β -carotene in the postpartum period on reproductive function of lactating Holstein cows in Florida.^a

Experiment	Time of year	Treatment	No. of cows	Pregnancy rate at first insemination	Pregnancy rate, 90-d postpartum
1	Hot	Control	128	9.4%	21.1%
		β -carotene	48	12.9%	35.4%*
2	Cool	Control	55	17.6%	33.3%
		β -carotene	52	12.9%	26.2%

^a From Aréchiga et al. (1998)

* P<0.05.

More recently, Garcia-Ispuerto et al. (2013) tested whether administration of 18 mg melatonin implants (9-12 implants per cow) beginning at 220 d of gestation to cows during the summer would affect postpartum reproduction. Melatonin has antioxidant properties in the follicle (Tamura et al., 2013). Cows treated with melatonin had reduced interval to conception and decreased incidence of cows experiencing > 3 breedings per conception (Table 2). It is not clear whether the effect of melatonin is due to blocking effects of heat stress on the oocyte and embryo or whether it acts through some effect independent of heat stress. Results are promising but experiments should be repeated in both hot and cool seasons.

Table 2. Effect of administration of melatonin implants at 220 d of gestation on postpartum reproduction in the summer in Spain.^a

Treatment	No. of cows	Cows with > 3 AI/conception	Interval from calving to conception, days
Control	67	28.4%	123
Melatonin	72	12.5%**	103*

^a From Garcia-Ispuerto et al., 2013

* P<0.05

** P< 0.01

Use of Hormones to Bypass Effects of Heat Stress on Reproduction

Protocols for timed artificial insemination (AI) can completely bypass problems associated with detecting estrus during heat stress because timing of ovulation is synchronized and insemination can be implemented at a fixed time without the need for estrus detection. What timed AI does not do is reverse damage to the oocyte or embryo caused by heat stress. Implementation of timed AI programs during heat stress can increase the rate at which cows get pregnant after calving because it increases the number of eligible cows that are inseminated. This is despite the fact that actual conception rates are not improved.

The effectiveness of timed AI during heat stress can be examined by looking at results from two experiments conducted in the summer (Table 3). The first experiment, that of Aréchiga et al. (1998), was conducted in south Florida, where the magnitude of heat stress is very high. Cows were assigned to either be bred at first detected estrus after the voluntary waiting period of 70 days or were subjected to the Ovsynch timed AI procedure to be inseminated at 70 d after calving. Implementation of timed AI reduced the interval from calving for first service by 10 d. There was no difference in the proportion of cows that became pregnant after first insemination between treatments. Moreover, few animals became pregnant, undoubtedly because of the high degree of heat stress. Nonetheless, more cows were pregnant by 90 d postpartum in the timed AI group, presumably because more cows had been inseminated.

Similar results were obtained in Kansas by Cartmill et al. (2001). In this experiment, one group of cows were subjected to an estrous synchronization regimen (Select Synch) whereas another group was inseminated using the Ovsynch protocol. The overall level of fertility was higher than for the Florida study. Like for the Florida experiment, timed AI did not increase the percent of cows pregnant at first insemination after calving. However, more cows were inseminated within 7 d after the TAI treatment than after the estrous synchronization treatment so the percent of cows pregnant at 90 d postpartum was almost twice as high for the timed AI group (Table 3).

Table 3. Effectiveness of timed insemination protocols for increasing pregnancy rates of lactating Holsteins when implemented during periods of heat stress in Florida (Experiment 1) and Kansas (Experiment 2).

Experiment ^a	Treatment ^b	No. of cows	Interval, calving to first service, days	Cows bred within 7 d after PGF	Pregnancy rate at first service	Pregnancy rate at 90-d postpartum
1	Estrus	184	82.4	--	12.5%	9.8%
	TAI	169	72.4***	--	13.6%	16.6%*
2	SS	128	--	57%	32.0%	17.9%
	TAI	207	---	100%	33.3%	33.3%**

^a Experiment 1: Aréchiga et al. (1998); Experiment 2, Cartmill et al. (2001).

^b Estrus = breeding at each observed estrus beginning at Day 70 postpartum; TAI = timed artificial insemination followed by breeding at all observed estrous periods thereafter; SS=Select Synch; GnRH followed by prostaglandin F-2 α (PGF) 7 d later and breeding at detected estrus for the next 21 d.

* P<0.05 **P<0.01 *** P<0.001

While timed AI can increase the number of cows inseminated, it cannot increase fertility during the summer. Unfortunately, no pharmacological treatment has been identified that can consistently increase pregnancy rate per AI during heat stress (see Hansen, 2011 for review). Among the treatments that have been examined are treatment with human chorionic gonadotropin at Day 5 of the estrous cycle to increase circulating progesterone concentrations, treatment with bovine somatotropin to increase secretion of the embryoprotective molecule insulin-like growth factor-1 and treatment with gonadotropin releasing hormone to extend lifespan of the corpus luteum. The ineffectiveness of hormonal treatments is probably related to the broad period of time in which the oocyte and early embryo are susceptible to disruption by heat shock. Treatments that might reverse effects of heat stress at one physiological window cannot reverse effects at others. Consider, for example, use of chorionic gonadotropin to increase output of progesterone by the corpus luteum. Such a treatment might be effective at reversing effect of heat stress on circulating progesterone concentrations after ovulation (Wolfenson et al., 2000) but this effect would not improve fertility in a cow whose oocyte was already damaged by heat stress occurring at some time before ovulation.

Embryo transfer to bypass death of oocytes and embryos caused by heat stress

Embryo transfer was developed as a tool to increase the number of offspring from genetically-superior females. This technology can also be used for improving fertility during heat stress. In fact, it remains the best method available for increasing pregnancy rate in lactating cows exposed to heat stress. Indeed, it has been shown repeatedly that embryo transfer can alleviate the reduction in fertility caused by heat stress (Figure 1).

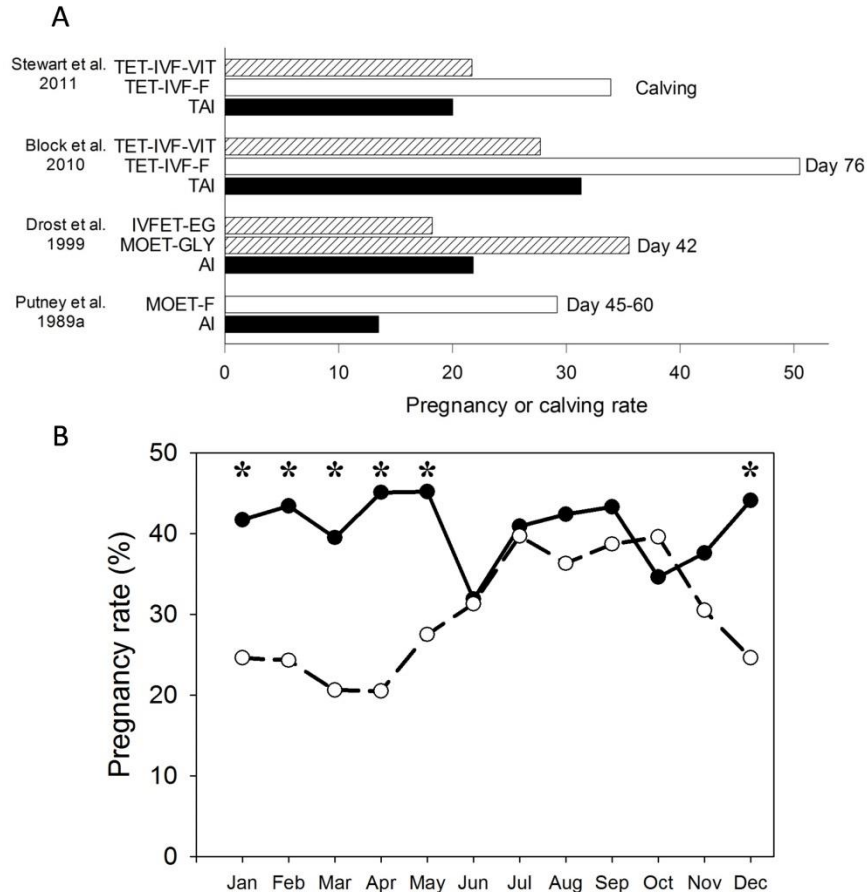


Figure 1. Enhancement of pregnancy rates during heat stress using embryo transfer. Data in Panel A represent results from various experiments in the summer in Florida. Abbreviations are as follows: AI: artificial insemination; EG, frozen in ethylene glycol; F, fresh; Gly, frozen in glycerol; IVFET, embryo transfer with an in vitro produced embryo; MOET, multiple ovulation embryo transfer; TAI, timed artificial insemination; TET-IVF, timed embryo transfer with an in vitro produced embryo; VIT, vitrified. The numbers in the graph represent the day of gestation at which pregnancy diagnosis was carried out. Panel B represents data from a commercial dairy in Brazil in which cows were either inseminated or received an embryo produced by superovulation (Rodrigues et al., 2004). Asterisks represent months in which pregnancy rate was different between AI and ET. The figure is reproduced from Hansen (2013).

Fertility is low in the summer largely because of damage to the growing follicle, oocyte and embryo caused by exposure to maternal hyperthermia (elevated body temperature). The oocyte can be damaged by heat stress as early as 105 days before ovulation (Torres-Júnior et al., 2008) and remains sensitive to heat stress on the day of ovulation (Putney et al., 1989b). The early embryo, too, can be damaged by heat stress but soon acquires biochemical mechanisms that protect it elevated temperature (Hansen, 2013). Thus, heat stress at Day 1 after estrus reduced embryonic development but heat stress at Days 3, 5, and 7 had no effect (Ealy et al., 1993).

In embryo transfer protocols, the only embryos typically transferred are those that have developed develop. Thus, the embryo that is transferred into a heat-stressed recipient has, for one reason or another, escaped

effects of heat stress. In addition, embryos have become resistant to heat stress by the time they reach the stage of development where they are ready to be transferred into a recipient (the morula or blastocyst stage). Thus, it is unlikely that maternal hyperthermia will kill an embryo after Day 7 of pregnancy than an embryo in the first day or two of life.

Embryos can be produced using either superovulation or in vitro fertilization. For superovulation, cows are injected with follicle stimulating hormone to cause the growth and ovulation of multiple follicles. For in vitro fertilization, oocytes are harvested either from growing follicles using transvaginal, ultrasound guided aspiration (called oocyte pickup or OPU) or from ovaries recovered at slaughter or ovariectomy. Oocytes are then fertilized with sperm and the resultant embryos allowed to develop in the laboratory until transferred into recipients. Embryos produced by superovulation are superior to those produced in vitro in terms of ability to establish pregnancy after transfer and survive cryopreservation for long-term storage. As shown in Figure 1, transfer of superovulated embryos improves fertility during heat stress regardless of whether embryos are cryopreserved or transferred fresh. However, the poor survival of in vitro produced embryos to cryopreservation means that transfer of in vitro produced embryos improved fertility only when embryos were transferred without cryopreservation.

Despite the problems caused by poor embryo freezability, in vitro production systems are superior to superovulation in terms of the maximum number of embryos that can be produced from a cow per year. In addition, sexed semen can be used very efficiently for in vitro fertilization since one straw can be used to inseminate dozens of oocytes. Production of in vitro produced embryos using oocytes collected at a slaughterhouse is much less expensive than production of embryos by superovulation or by in vitro fertilization of oocytes harvested using OPU.

The decision as to whether to use embryo transfer during the summer depends on the magnitude of the reduction in fertility caused by heat stress, the degree of improvement in fertility caused by embryo transfer and the cost of the embryo available for transfer. Ribeiro et al. (2012) estimated the cost to produce a female pregnancy in lactating cows as a function of the pregnancy rate per AI or embryo transfer. As an example, consider the case where pregnancy rate in the summer is 15% to AI using conventional semen and 25% using embryo transfer with sexed semen. In this scenario, embryo transfer would be profitable. It would cost \$1157 to produce a female pregnancy using timed AI, \$1042 to produce an embryo using an oocyte harvested by ultrasound, and \$820 to produce an embryo using an oocyte recovered from a slaughterhouse ovary.

Embryo transfer will become more profitable during heat stress as improvements in the process increase the competence of the embryo to establish pregnancy and survive cryopreservation. In addition, cost advantages of using embryos derived from oocytes recovered from the slaughterhouse should not be overlooked. Embryos of high genetic merit can be produced by using elite bulls because one straw of semen can produce dozens of embryos. Improvements in cryopreservation will also make embryo biopsy for genotyping more practical.

References

Aréchiga CF, Staples CR, McDowell LR, Hansen PJ. Effects of timed insemination and supplemental β -carotene on reproduction and milk yield of dairy cows under heat stress. *J Dairy Sci* 1998; 81:390-402.

Badinga L, Collier RJ, Thatcher WW, Wilcox CJ. Effects of climatic and management factors on conception rate of dairy cattle in subtropical environment. *J Dairy Sci* 1985; 68:78-85.

Berman, A., Folman, Y., Kaim, M., Mamen, M., Herz, Z., Wolfenson, D., Arieli, A., & Graber, Y. (1985) Upper critical temperatures and forced ventilation effects for high-yielding dairy cows in a subtropical climate. *J Dairy Sci* 1985; 68:1488-1495.

Cartmill JA, El-Zarkouny SZ, Hensley BA, Rozell TG, Smith JF, Stevenson JS. An alternative AI breeding protocol for dairy cows exposed to elevated ambient temperatures before or after calving or both. *J Dairy Sci* 2001; 84:799-806.

de Castro e Paula LA, Hansen PJ. Modification of actions of heat shock on development and apoptosis of cultured preimplantation bovine embryos by oxygen concentration and dithiothreitol. *Mol Reprod Dev* 2008; 75:1338-1350.

Dikmen S, Alava E, Pontes E, Fear JM, Dikmen BY, Olson TA, Hansen PJ. Differences in thermoregulatory ability between slick-haired and wild-type lactating Holstein cows in response to acute heat stress. *J Dairy Sci* 2008; 91:3395-3402.

Dikmen S, Hansen PJ. Is the temperature-humidity index the best indicator of heat stress in lactating dairy cows in a subtropical environment? *J Dairy Sci* 2009; 92: 109-116.

Drost, M, Ambrose JD, Thatcher M-J, Cantrell CK, Wolfsdorf KE, Hasler JF, Thatcher WW. Conception rates after artificial insemination or embryo transfer in lactating dairy cows during summer in Florida. *Theriogenology* 1999; 52:1161-1167.

Ealy AD, Drost M, Hansen PJ. 1993. Developmental changes in embryonic resistance to adverse effects of maternal heat stress in cows. *J Dairy Sci* 1993; 76:2899-2905.

Ealy AD, Howell JL, Monterroso VH, Aréchiga CF, Hansen PJ. Developmental changes in sensitivity of bovine embryos to heat shock and use of antioxidants as thermoprotectants. *J Anim Sci* 1995; 73:1401-1407.

Flamenbaum I, Ezra E. Cooling cows in summer almost eliminates seasonality in milk production and fertility. In: *The Dairy Industry in Israel 2006*. (ed. D Hojman, Malul Y, Avrech T), Israel Cattle Breeders Assn and Israel Dairy Board, Israel, 2006, pp 23-25.

Garcia-Ispuerto I, Abdelfatah A, López-Gatius F. Melatonin treatment at dry-off improves reproductive performance postpartum in high-producing dairy cows under heat stress conditions. *Reprod Domest Anim* 2013; 48:577-583.

Gaughan JB, Mader TL, Holt SM, Sullivan ML, Hahn GL. Assessing the heat tolerance of 17 beef cattle genotypes. *Int J Biometeorol* 2010; 54:617-627.

Gilad E, Meidan R, Berman A, Graber Y, Wolfenson D. Effect of tonic and GnRH-induced gonadotrophin secretion in relation to concentration of oestradiol in plasma of cyclic cows. *J Reprod Fertil* 1993; 99: 315-321.

Hansen PJ. Managing reproduction during heat stress in dairy cows. In: Risco CA, Melendez Retamel P, *Dairy Production Medicine*, Wiley-Blackwell, Chicester, United Kingdom, 2011, pp. 153-163.

Hansen PJ. 2013. Cellular and molecular basis of therapies to ameliorate effects of heat stress on embryonic development in cattle. *Anim Reprod* 2013; 10: 322-333.

Hansen PJ, Aréchiga CF. Strategies for managing reproduction in the heat-stressed dairy cow. *J Anim Sci* 1999; 77 Suppl 2:36-50.

López-Gatius, F, Santolaria P, Mundet I, Yániz JL. Walking activity at estrus and subsequent fertility in dairy cows. *Theriogenology* 2005; 63:1419-1429.

Mitlöhner FM, Galyean ML, McGlone JJ. Shade effects on performance, carcass traits, physiology, and behavior of heat-stressed feedlot heifers. *J Anim Sci* 2002; 80:2043-2050.

Nabenishi H, Takagi S, Kamata H, Nishimoto T, Morita T, Ashizawa K, Tsuzuki Y. The role of mitochondrial transition pores on bovine oocyte competence after heat stress, as determined by effects of cyclosporin A. *Mol Reprod Dev* 2012; 79:31-40.

Paula-Lopes FF, Al-Katanani YM, Majewski AC, McDowell LR, Hansen PJ. Manipulation of antioxidant status fails to improve fertility of lactating cows or survival of heat-shocked embryos. *J Dairy Sci* 2003; 86:2343-2351.

Putney DJ, Drost M, Thatcher WW. Influence of summer heat stress on pregnancy rates of lactating dairy cattle following embryo transfer or artificial insemination. *Theriogenology* 1989a; 31:765-778.

Putney DJ, Mullins S, Thatcher WW, Drost M, Gross TS. Embryonic development in superovulated dairy cattle exposed to elevated ambient temperatures between the onset of estrus and insemination. *Anim Reprod Sci*, 1989b; 19:37-51.

Ribeiro ES, Galvão KN, Thatcher WW, Santos JEP. Economic aspects of applying reproductive technologies to dairy herds. *Anim. Reprod* 2012; 9:370-387.

Rodrigues CA, Ayres H, Reis EL, Nichi M, Bo GA, Baruselli PS. Artificial insemination and embryo transfer pregnancy rates in high production Holstein breedings under tropical conditions. *Proc 15th Int Congr Anim Reprod* 2004; 2: 396 (abstr).

Sakatani M, Kobayashi S, Takahashi M. Effects of heat shock on in vitro development and intracellular oxidative state of bovine preimplantation embryos. *Mol Reprod Dev* 2004; 67:77-82.

Sakatani M, Suda I, Oki T, Kobayashi S, Kobayashi S, Takahashi M. Effects of purple sweet potato anthocyanins on development and intracellular redox status of bovine preimplantation embryos exposed to heat shock. *J Reprod Dev* 2007; 53:605-614.

Stewart BM, Block J, Morelli P, Navarette AE, Amstalden M, Bonilla L, Hansen PJ, Bilby TR. Efficacy of embryo transfer in lactating dairy cows during summer using fresh or vitrified embryos produced in vitro with sex-sorted semen. *J Dairy Sci* 2011; 94:3437-3445.

Tamura H, Takasaki A, Taketani T, Tanabe M, Kizuka F, Lee L, Tamura I, Maekawa R, Asada H, Yamagata Y, Sugino N. Melatonin as a free radical scavenger in the ovarian follicle. *Endocrine J* 2013; 60:1-13.

Thatcher WW, Collier RJ. Effects of climate on bovine reproduction. In: *Current Therapy in Theriogenology* 2. (Morrow DA, ed), 1986; pp. 301-309. W. B. Saunders, Philadelphia.

Torres-Júnior JR de S, Pires M de FA Pires, de Sá WF, Ferreira A de M, Viana JH, Camargo LS, Ramos AA, Folhadella IM, Polisseni J, de Freitas C, Clemente CA, de Sá Filho MF, Paula-Lopes FF, Baruselli PS. Effect of maternal heat-stress on follicular growth and oocyte competence in *Bos indicus* cattle. *Theriogenology*, 2008; 69:155-166.

Wolfenson D, Roth Z, Meidan R. Impaired reproduction in heat-stressed cattle: basic and applied aspects. *Anim Reprod Sci* 2000; 60-61:535-547.